

DELIVERY DYNAMICS AND DESTINATION OF GENE VECTOR INSTILLATIONS IN LIVE MOUSE AIRWAYS

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BACKGROUND: Most groups use the mouse nose as an *in vivo* model site for developing gene treatments for CF airway disease. Reliable dosing of an airway pre-treatment followed by a lentiviral vector is essential for the success of gene therapy. Despite standardised delivery techniques we see variability in reporter-gene histological assessments and electrophysiological measurements. The aim of this experiment was to use synchrotron X-ray imaging at SPring-8 to accurately determine the fate of fluids delivered into live mouse nasal airways and the trachea.

METHODS: Newborn C57BL/6J mice ($n = 6$ nasal & $n = 4$ tracheal) were imaged on the BL20B2 beamline at the SPring-8 synchrotron (Japan). For the nasal study images were captured at 1 Hz. After 1 min of baseline a 4 μ l sample of iodine-based contrast fluid (airway pre-treatment surrogate) was delivered over 10 sec. Imaging continued for a further 10 min. Fluid motion was revealed using a background subtraction method. For the tracheal study mice were intubated and ventilated at 20 breaths/min with 1 ml/min image acquisition rate per breath. At baseline a 15 μ l bolus (airway pre-treatment surrogate) was delivered over 30 sec. Following 1 min of data collection an additional 15 μ l bolus was delivered over 3.6 sec. Image capture continued for 10 min. Frame differencing was used to reveal fluid motion.

NASAL AIRWAY RESULTS:

- The 4 μ l dose (F1) was retained on the treated side, supporting the strength of gene expression seen in nasal ciliated epithelium, validating the use of a 4 μ l dose to retain an untreated nasal airway as a whole.
- Confirms why gene expression is limited to the treated nostril, as it extends into the tubular nasopharynx.
- Though the dose also distributes into the lateral transitional/olfactory region, expression from the gene vector was restricted to respiratory epithelium in our gene delivery system.
- The 20 μ l dose (F2) overwhelms the 'holding capacity' of the dosed nostril, with some fluid continuing into the trachea and then very little deeper lung airways.
- Supports our finding of long-term lung expression using gene vectors delivered via nose and suggests smaller vector doses may produce the same effect in the lungs.
- Supports the specificity of gene delivery to only certain general lung areas reached by the 4 μ l dose, despite demonstrating the wide reach of the 20 μ l dose.



CONCLUSION: Synchrotron imaging can help explain the mechanisms underlying published outcomes from our gene-transduction protocols in mouse nasal airways. Our findings suggest the need for, and permit, much greater attention to dosing technique – orientation, volume, speed – and enable improvements in dosing technique design.

ACKNOWLEDGEMENTS: Funding from NHMRC, Australian Synchrotron ISAP Program, JASRI and www.Cure4CF.org. We also thank Andreas Foures, Naoto Yagi and Kentaro Uesugi.